

# Role of Hydrogen Sulfide in Acute Lung Injury and Acute Respiratory Distress Syndrome

Huili Zhang<sup>1</sup> and Madhav Bhatia<sup>\*2</sup>

<sup>1</sup>Department of Cardiology, Shanghai Ninth People's Hospital, Shanghai JiaoTong University School of Medicine, China

<sup>2</sup>Department of Pharmacology, National University of Singapore, Singapore

**Abstract:** Accumulating evidence has suggested that hydrogen sulfide (H<sub>2</sub>S) naturally occurs during cysteine metabolism in many types of mammalian cells. Since H<sub>2</sub>S exhibits vasodilator activity and plays an important role in nervous system and inflammatory diseases, it is currently considered to be the third gaseous mediator. Recently, more and more attention has been paid to the biological functions of H<sub>2</sub>S in acute lung injury (ALI) and/or acute respiratory distress syndrome (ARDS). In various animal models of lung injury, H<sub>2</sub>S has been demonstrated to contribute to the development and progression of lung inflammation and injury. Regulating the endogenous level of H<sub>2</sub>S is possible to protect animals against lung injury. H<sub>2</sub>S may exert its effect on ALI/ARDS by modulating leukocyte activation. In addition, H<sub>2</sub>S may induce lung inflammation and injury *via* activating sensory nerves in lung and eliciting a neurogenic inflammatory response.

## INTRODUCTION

Hydrogen sulfide (H<sub>2</sub>S) has been recognized as a toxic gas with a characteristic odor of rotten eggs for nearly 300 years [1]. However, it is now evident that H<sub>2</sub>S is endogenously generated in many types of mammalian cells and tissues during cysteine metabolism in a reaction catalyzed by cystathionine-γ-lyase (CSE, EC 4.4.1.1) and/or cystathionine-β-synthetase (CBS, EC 4.2.1.22) [2-4]. Although endogenous H<sub>2</sub>S was detected in the brainstem at the end of 1980s, its biological role and specific cellular targets in nervous system was first identified a decade ago [5]. Now H<sub>2</sub>S is increasingly considered as the third gasotransmitter.

Although much less is known about the biological functions of H<sub>2</sub>S than about its two counterparts, nitric oxide (NO) and carbon monoxide (CO), the gas is suggested to fulfill a wide range of physiological and pathological functions. For instance, H<sub>2</sub>S opens K<sup>+</sup><sub>ATP</sub> channels in vascular smooth muscle cells, gastrointestinal smooth muscle cells, cardiomyocytes, neurons, and pancreatic β-cells, therefore regulates vascular tone, intestinal contractility, myocardial contractility, neurotransmission and insulin secretion [6-8]. In nervous system, H<sub>2</sub>S promotes hippocampal long-term potentiation (LTP) by enhancing the sensitivity of NMDA receptors to glutamate and plays a role in neurodegenerative diseases [6-8]. In addition, H<sub>2</sub>S may scavenge reactive nitrogen species (RNS), peroxynitrite (OONO<sup>-</sup>), oxygen free radicals and lipid peroxidations, resulting in cardiovascular protection and neuron protection [9-11].

Despite the vasodilator and atypical neuromodulator activity of H<sub>2</sub>S, it has recently been shown to play an important

role in the pathogenesis of inflammatory diseases and associated organ injury, such as acute pancreatitis, sepsis and endotoxemia [12-16]. Overproduction of endogenous H<sub>2</sub>S may exacerbate inflammatory response and lung injury in acute pancreatitis, sepsis and endotoxemia and inhibition of H<sub>2</sub>S formation may be a potential therapeutic approach in these diseases.

## Biosynthesis and Metabolism of H<sub>2</sub>S

It is well known that H<sub>2</sub>S can be generated by breaking down organic matters in certain bacteria and archaea. Interestingly, many types of mammalian cells can produce H<sub>2</sub>S. A substantial amount of H<sub>2</sub>S is found to be present both in circulation and in various tissues, such as liver, kidney, pancreas, brain, aorta etc [6-8]. For instance, the concentration of H<sub>2</sub>S in rat and human serum is reported to be 46 and 43.8 μM, respectively [13, 17]. The physiological level of H<sub>2</sub>S in central nervous system is approximately 50-160 μM [5, 18].

The majority of endogenous H<sub>2</sub>S generation is catalyzed by cystathionine-γ-lyase (CSE, EC 4.4.1.1) and/or cystathionine-β-synthetase (CBS, EC 4.2.1.22), which use L-cysteine as the main substrate [2-4]. These two enzymes are pyridoxal-5'-phosphate-dependent. Although CBS and CSE are widely expressed in cells and tissues, CSE is the predominant H<sub>2</sub>S-forming enzyme in cardiovascular system, liver, kidney and non-vascular smooth muscle cells whereas CBS is mainly distributed in nervous system. As the end product of CSE- and CBS-involved cysteine metabolism, H<sub>2</sub>S has a negative feedback on the activity of these two enzymes [19, 20]. In addition, a small proportion of endogenous H<sub>2</sub>S may be generated by non-enzymatic steps in erythrocytes [21].

H<sub>2</sub>S *in vivo* is metabolized by oxidation in mitochondria or by methylation in cytosol [6]. It can also be scavenged by methemoglobin or by metallo- or disulfide-containing molecules such as oxidized glutathione [6]. In addition, H<sub>2</sub>S is

\*Address correspondence to this author at the Cardiovascular Biology Research Programme, Department of Pharmacology, Centre for Life Sciences, National University of Singapore, 28 Medical Drive, #03-02, Singapore 117456; Tel: (65)-6516-8256; Fax: (65)-6775-7674; E-mail: mhatia@nus.edu.sg

excreted mainly by the kidney as free or conjugated sulfate [6].

### **Etiology and Pathogenesis of Acute Lung Injury (ALI) and Acute Respiratory Distress Syndrome (ARDS)**

ALI and its most severe manifestation, ARDS, is a clinical syndrome characterized by acute hypoxemic respiratory failure, bilateral pulmonary infiltrates on frontal chest radiograph consistent with edema, and normal cardiac filling pressures [22, 23]. ALI/ARDS may occur in patients of all ages from direct or indirect insults. Common direct pulmonary causes of ALI/ARDS include lung viral or bacterial infections, gastric aspiration, blunt thoracic trauma with lung contusion, meconium aspiration (infants), near-drowning, thoracic radiation, hyperoxia, and the inhalation of smoke or other toxicants. Common indirect (systemic) causes of ALI/ARDS include sepsis, closed space burn injury, hypovolemic shock, non-thoracic trauma, multiple transfusions, and pancreatitis. These direct or indirect insults induce pulmonary inflammation, damage the pulmonary capillary permeability and alveolar diffusion capacity, increase intrapulmonary shunt, leading to severe acute respiratory failure [23-28]. Furthermore, complex autocrine and paracrine interrelationships of cytokines and pro-inflammatory mediators initiate and amplify the inflammatory response in ALI/ARDS. Cellular responses including endothelial adhesion molecules expression and migration of PMNs, as well as humoral responses, such as lipid mediators, proteases, oxidants, growth factors, NO, neuropeptides, and nuclear factor- $\kappa$ B (NF- $\kappa$ B) also participate in the pathogenesis of ALI/ARDS [29].

### **Role of H<sub>2</sub>S in ALI/ARDS of Different Etiologies**

In the past few years, H<sub>2</sub>S has emerged as a novel and increasingly important mediator in inflammatory diseases. Its role in ALI/ARDS has also been investigated in different animal models and clinical cases. Although H<sub>2</sub>S may exert different effect in ALI/ARDS induced by different insults, it has suggested that endogenous H<sub>2</sub>S may participate in the pathogenesis of ALI/ARDS and H<sub>2</sub>S related therapy may be a potential therapeutic approach in this condition.

#### **Pro-Inflammatory**

##### ***Sepsis or Endotoxemia Induced Lung Injury***

Sepsis and its sequelae such as septic shock and multiple organ failure are common and serious medical problems in severely ill patients [30]. An overproduction of endogenous H<sub>2</sub>S in vascular tissues has been found in cecal ligation and puncture (CLP) induced septic shock or lipopolysaccharide (LPS) induced endotoxemic shock. The vascular levels of H<sub>2</sub>S may be associated with the hemodynamic collapse during shock [31]. Recently, it has been shown that generation of endogenous H<sub>2</sub>S is elevated in plasma, liver and kidney, during CLP induced sepsis as well as LPS induced endotoxemia [13, 15]. Plasma H<sub>2</sub>S concentration is also significantly increased in septic patients compared to healthy controls [13]. Furthermore, inhibition of H<sub>2</sub>S formation by administration with DL-propargylglycine (PAG), a CSE inhibitor not only suppresses the overproduction of pro-inflammatory cytokines and chemokines in sepsis, which is a key feature of systemic inflammation, but also attenuates acute lung injury caused by sepsis or endotoxemia [13-16]. Notably, the protec-

tive effect of PAG is independent of the improvement of hemodynamics, since it did not alter blood pressure [14]. In contrast, H<sub>2</sub>S donors aggravate inflammation and sepsis associated ALI [15]. The pro-inflammatory effect of H<sub>2</sub>S in sepsis may be due to the activation of ERK-NF- $\kappa$ B pathway [16, 32]. In addition, injection of NaHS to normal mice directly results in lung inflammation and inflammatory damage in a dose-dependent manner [33]. Administration with NaHS at a dose of 10 mg/kg but not 1 or 5 mg/kg, caused an obvious lung injury as evidence by lung MPO activity and histological changes in lung sections [33].

##### ***Acute Pancreatitis Induced Lung Injury***

Since both CBS and CSE, two H<sub>2</sub>S forming enzymes, are highly expressed in pancreatic acinar cells, some studies investigated the potential role of H<sub>2</sub>S in acute pancreatitis [34]. Induction of acute pancreatitis in mice by caerulein increased plasma H<sub>2</sub>S level. In pancreatic acinar cells stimulated by caerulein, the production of H<sub>2</sub>S and CSE mRNA were significantly elevated, while the expression of CBS was reduced, suggesting that CSE is the main H<sub>2</sub>S forming enzyme in caerulein induced acute pancreatitis [34]. Pretreatment or posttreatment with PAG to inhibit the activity of CSE not only attenuated the inflammatory response (MPO activity and chemokine expression) in pancreas and lung but also mitigated the severity of pancreatitis associated lung injury [35, 36].

#### **Anti-Inflammatory**

##### ***Burn and Smoke Inhalation Induced Lung Injury***

In a murine model of acute lung injury induced by combined burn and smoke inhalation, intraperitoneal administration of NaHS, an H<sub>2</sub>S donor alleviated lung injury, as evidenced by a promising improvement in survival rate and lung histological conditions [37]. H<sub>2</sub>S may exert the protective effect *via* up-regulating the level of tissue anti-inflammatory cytokines (IL-10) and reducing the generation of pro-inflammatory cytokines (IL-1 $\beta$ ). In this model, H<sub>2</sub>S can also rescue smoke exposed lung from oxidative stress and therefore improve the outcome of ALI.

##### ***Oleic Acid Induced Lung Injury***

In oleic acid induced ALI, a significant reduction in H<sub>2</sub>S levels in plasma and lung tissues was observed [38]. However, administration of H<sub>2</sub>S donors in rats treated with oleic acid down-regulated the production of pro-inflammatory IL-6 and IL-8 in lung but increased the level of anti-inflammatory IL-10 in plasma and lung, thus alleviating lung edema, PMNs infiltration in lung and severity of lung injury.

##### ***Application of H<sub>2</sub>S - Releasing NSAIDs in ALI/ARDS***

S-diclofenac (ACS 15) is a type of H<sub>2</sub>S-releasing diclofenac, which comprises a H<sub>2</sub>S-releasing dithiol-thione moiety attached by an ester linkage to diclofenac. Prophylactic or therapeutic administration of ACS 15 significantly attenuated lung inflammation and the severity of lung injury induced by acute pancreatitis but had no significant effect on pancreatic damage, suggesting the usefulness of H<sub>2</sub>S-releasing nonsteroidal anti-inflammatory drugs as a potential treatment for pancreatitis-associated ALI/ARDS [39]. ACS15 also exhibits enhanced anti-inflammatory and protective effect as compared to the parent drug in a rat model of

endotoxemia and associated lung injury [40]. It is the H<sub>2</sub>S released from ACS 15 that may intensify the anti-inflammatory activity of ACS 15 by inhibiting the DNA binding activity of nuclear transcriptional factors (AP-1 and NF-κB) and consequent production of inflammation related genes [40].

### Potential Mechanisms of the Role of H<sub>2</sub>S in ALI/ARDS

#### Regulation of Leukocyte Activity

Recent studies have revealed that H<sub>2</sub>S may up-regulate the inflammatory response *via* stimulation of immune cells. It has been shown that reactive oxygen species from activated neutrophils converted H<sub>2</sub>S to sulfite, which may up-regulate leukocyte adhesion and neutrophil functions [41-43]. The reaction was suppressed by the NADPH oxidase inhibitor and was accelerated by the addition of NaHS. Serum levels of H<sub>2</sub>S and sulfite was found to be elevated in LPS injected rats. These data imply that oxidative stress dependent conversion of H<sub>2</sub>S to sulfite by activated neutrophils may enhance non-specific host defense in various inflammatory diseases such as pneumonia [43, 44]. Most recently, Zhi *et al.* reported that H<sub>2</sub>S dose-dependently activated human monocytes with up-regulating the production of pro-inflammatory cytokines, at least partially *via* the activation of extracellular signal-regulated kinase (ERK)-NF-κB signaling pathway [45]. Pretreatment with NF-κB inhibitor or MEK antagonist significantly inhibited H<sub>2</sub>S induced activation of NF-κB and secretion of pro-inflammatory cytokines [45]. In addition, H<sub>2</sub>S *in vitro* provoked the short-term survival of granulocyte *via* inhibition of caspase-3 cleavage and p38 mitogen-activated protein kinase (MAPK) activation and therefore contributed to the bactericidal activity of neutrophils [46].

In contrast, some *in vitro* studies performed in macrophages propose the anti-inflammatory role of H<sub>2</sub>S. For instance, in cultured murine RAW264.7 macrophages, H<sub>2</sub>S down-regulated LPS induced iNOS expression, NO production and NF-κB activation. This anti-inflammatory effect may be mediated by H<sub>2</sub>S induced activation of ERK and consequent up-regulation of HO-1 expression and CO production, which exerts inhibitory effect on LPS-induced activation of NF-κB and resulted in a reduction in NO synthesis [47]. In addition, Hu *et al.* observed the effect of H<sub>2</sub>S on LPS induced inflammation in both primary and cultured microglia and immortalized murine BV-12 microglia cells [48]. They found that both endogenous and exogenous H<sub>2</sub>S significantly reduced LPS-induced NO production and TNF-α secretion in microglia. The effect of H<sub>2</sub>S was mediated by inhibition of LPS induced activation of p38 MAPK. However, the anti-inflammatory role of H<sub>2</sub>S reported in this study is merely limited to CNS-derived glia cells, in which CBS but not CSE is the primary H<sub>2</sub>S-producing enzyme.

#### Regulation of Leukocyte Trafficking

Recently, one *in vivo* experiment indicates that H<sub>2</sub>S may contribute to leukocyte-endothelial interaction, such as leukocyte rolling and adhesion, as well as promote PMN infiltration (MPO activity) into inflamed tissues during polymicrobial sepsis or endotoxemia [13, 15]. H<sub>2</sub>S may provoke leukocyte migration by the activation of NF-κB and consequent up-regulation of the expression of adhesion molecules

[49]. Furthermore, administration of NaHS in normal mice caused a significant increase in DNA binding activity of NF-κB and production of adhesion molecules [16, 49]. In addition, H<sub>2</sub>S has been found to up-regulate the expression of CXCR2 in PMNs and therefore facilitate MIP-2 directed migration of PMNs [49].

On the other hand, some investigators reported that H<sub>2</sub>S donors attenuated NSAIDs-induced gastric granulocyte infiltration and leukocyte adherence in mesenteric venules likely *via* opening K<sub>ATP</sub> channels [50, 51]. Leukocyte infiltration in the air pouch in response to carrageenan was also suppressed by NaHS [50, 51]. This effect was reversed by either inhibition of H<sub>2</sub>S formation or pretreatment with K<sub>ATP</sub> channel blocker. In addition, H<sub>2</sub>S also inhibited fMLP-induced leukocyte adherence to the mesenteric microcirculation [51]. The inconsistency in the role of H<sub>2</sub>S in regulating leukocyte migration may be due to the different animal models and different doses of H<sub>2</sub>S donors used.

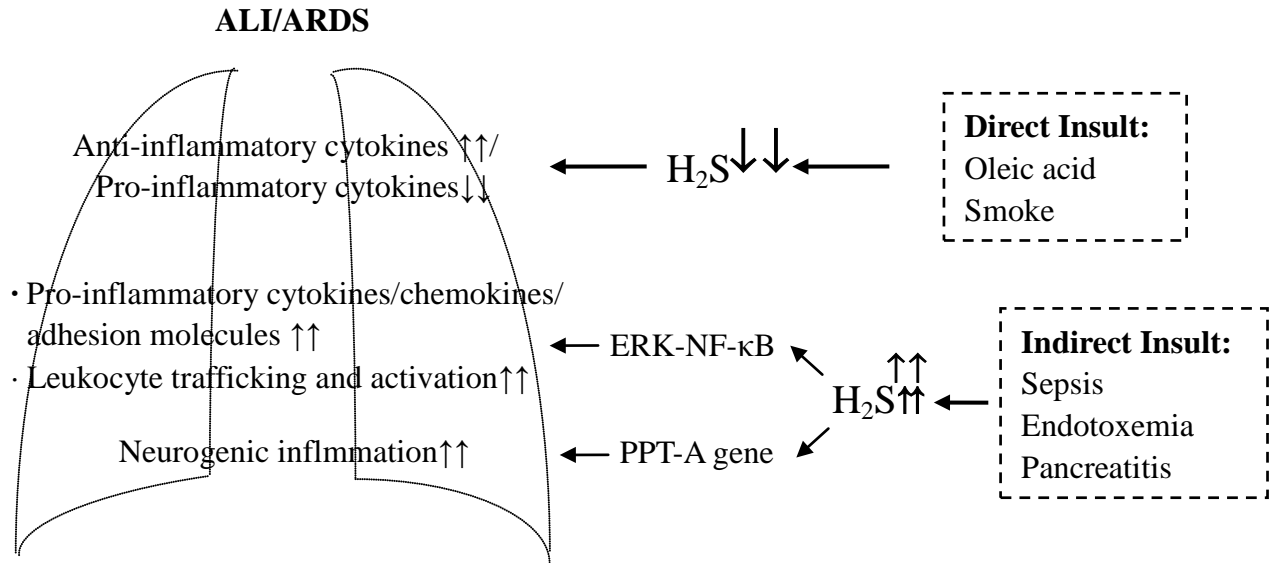
#### Neurogenic Inflammation

Activation of afferent sensory nerve causes the release of bioactive substances, such as substance P, from never terminal, leading to vasodilation, edema and other manifestation of inflammation. This phenomenon is called "neurogenic inflammation" [52-54]. The association between H<sub>2</sub>S and substance P in lung inflammation and injury has been investigated both *in vitro* and *in vivo*. Trevisani *et al.* found that NaHS not only provoked the release of substance P and CGRP from the sensory nerve terminals in isolated guinea pig airways but also produced a concentration-dependent contractile response [55]. Recently, one study performed by our group found that i.p. administration of NaHS in normal mice caused a significant rise in the circulatory level of substance P in a dose-and time-dependent manner, coupled with obvious lung inflammation [33]. Genetic deletion of *preprotachykinin-A (PPT-A)* gene, the precursor gene for substance P, or depletion of substance P from sensory neurons by capsaicin or pretreatment with capsazepine, an antagonist of the transient receptor potential vanilloid-1 (TRPV-1), abolished the inflammatory role of H<sub>2</sub>S and therefore protected normal mice against H<sub>2</sub>S-induced lung inflammation. This study shows for the first time that H<sub>2</sub>S may induce neurogenic inflammation and lung injury even without other noxious stimuli.

In sepsis and associated lung injury, H<sub>2</sub>S may exert its pro-inflammatory effect *via* up-regulating the production of substance P. Inhibition of the activation of substance P by genetic depletion of *PPTA* gene prevented NaHS from exacerbating inflammatory response and aggravating lung injury in sepsis [56]. Consistent findings were obtained in a murine model of acute pancreatitis. Inhibition of H<sub>2</sub>S formation by PAG significantly decreased the expression of substance P in lung and pancreas in caerulein-induced acute pancreatitis and associated lung injury [57]. Taken together, both *in vitro* and *in vivo* studies suggest that neurogenic inflammation may mediate the pro-inflammatory role of H<sub>2</sub>S in ALI/ARDS.

### CONCLUSION AND PERSPECTIVES FOR THE FUTURE

H<sub>2</sub>S is now considered to play a key role in inflammation. However, its role in ALI/ARDS is still far from clear,



**Fig. (1).** A schematic summary of the evidence for the potential role of  $H_2S$  in ALI/ARDS. ALI/ARDS induced by direct or indirect insults may alter the circulatory level of  $H_2S$ .  $H_2S$  released from vascular smooth cells, liver, pancreas or lung (?) may regulate the production of inflammatory mediators (cytokines, chemokines and adhesion molecules), and leukocyte activation in lung by activating nuclear NF- $\kappa$ B, therefore leading to ALI/ARDS.  $H_2S$  may also facilitate inflammatory manifestations in lung, such as edema and plasma protein extravasation, by promoting neurogenic inflammation *via* upregulating the expression of PPT-A or NK1R gene.

and Fig. (1) summarized the relevant findings obtained to date. The apparent discrepancy in the role of  $H_2S$  in ALI/ARDS may be due to the different animal models and different doses of  $H_2S$  donors used. The protective effect of  $H_2S$  in ALI has been reported in some models of ALI caused by direct insults (smoke inhalation) whereas the opposite findings are usually obtained in ALI caused by indirect or systemic insults (e.g. sepsis, pancreatitis). Furthermore, high dose of  $H_2S$  donor tends to aggravate lung inflammation and injury while the low dose (or slow release) of  $H_2S$ , which approximates the physiological concentration but below that observed following inflammation, has been reported to mitigate ALI.

Further research is needed in the following directions: (1) to further characterize the precise mechanisms by which  $H_2S$  contributes to the development and progression of ALI/ARDS, including its interaction with other molecules; (2) to investigate whether  $H_2S$  participate in the pathogenesis of ALI/ARDS in humans and whether these results can be translated into the clinic; (3) to explore whether inhibition of  $H_2S$  biosynthesis provide a novel and potential forward for the development of treatment for ALI/ARDS. Although research in these directions is still in its infancy, the role of  $H_2S$  in ALI/ARDS is likely to be a complex one.

## REFERENCES

- [1] Beauchamp RO, Bus JS, Popp JA, Boreiko CJ, Andjelkovich DA. A critical review of the literature on hydrogen sulfide toxicity. *CRC Crit Rev Toxicol* 1984; 13: 25-97.
- [2] Bukovska G, Kery V, Kraus JP. Expression of human cystathionine  $\beta$ -synthase in *Escherichia coli*: purification and characterization. *Protein Exp Purif* 1994; 5: 442-8.
- [3] Erickson PF, Maxwell IH, Su LJ, Baumann M, Glode LM. Sequence of cDNA for rat cystathionine  $\gamma$ -lyase and comparison of deduced amino acid sequence with related *Escherichia coli* enzymes. *Biochem J* 1990; 269: 335-40.
- [4] Stipanuk MH, Beck PW. Characterization of the enzymic capacity for cysteine desulphhydration in liver and kidney of the rat. *Biochem J* 1982; 206: 267-77.
- [5] Abe K, Kimura H. The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 1996; 16: 1066-71.
- [6] Wang R. Two's company, three's a crowd: can  $H_2S$  be the third endogenous gaseous transmitter? *FASEB J* 2002; 16: 1792-8.
- [7] Moore PK, Bhatia M, Moochhala SM. Hydrogen sulfide: from the smell of the past to the mediator of the future? *Trends Pharmacol Sci* 2003; 24: 609-11.
- [8] Lowicka E, Beltowski J. Hydrogen sulfide ( $H_2S$ ) - the third gas of interest for pharmacologists. *Pharmacol Rep* 2007; 59: 4-24.
- [9] Geng B, Chang L, Pan C, *et al.* Endogenous hydrogen sulfide regulation of myocardial injury induced by isoproterenol. *Biochem Biophys Res Commun* 2004; 318: 756-63.
- [10] Kimura Y, Kimura H. Hydrogen sulfide protects neurons from oxidative stress. *FASEB J* 2004; 18: 1165-7.
- [11] Whiteman M, Armstrong JS, Chu SH, *et al.* The novel neuromodulator hydrogen sulfide: an endogenous peroxynitrite 'scavenger'? *J Neurochem* 2004; 90: 765-8.
- [12] Bhatia M, Wong FL, Fu D, Lau HY, Moochhala SM, Moore PK. Role of hydrogen sulfide in acute pancreatitis and associated lung injury. *FASEB J* 2005; 19: 623-5.
- [13] Li L, Bhatia M, Zhu YZ, *et al.* Hydrogen sulfide is a novel mediator of lipopolysaccharide-induced inflammation in the mouse. *FASEB J* 2005; 19: 1196-8.
- [14] Collin M, Anuar FB, Murch O, Bhatia M, Moore PK, Thiernemann C. Inhibition of endogenous hydrogen sulfide formation reduces the organ injury caused by endotoxemia. *Br J Pharmacol* 2005; 146: 498-505.
- [15] Zhang H, Zhi L, Moore PK, Bhatia M. Role of hydrogen sulfide in cecal ligation and puncture-induced sepsis in the mouse. *Am J Physiol Lung Cell Mol Physiol* 2006; 290: L1193-201.
- [16] Zhang H, Zhi L, Moochhala S, Moore PK, Bhatia M. Hydrogen sulfide acts as an inflammatory mediator in cecal ligation and puncture-induced sepsis in mice by upregulating the production of cytokines and chemokines *via* NF- $\kappa$ B. *Am J Physiol Lung Cell Mol Physiol* 2007; 292: L960-71.
- [17] Zhao W, Zhang J, Lu Y, Wang R. The vasorelaxant effect of  $H_2S$  as a novel endogenous gaseous KATP channel opener. *EMBO J* 2001; 20: 6008-16.
- [18] Hosoki R, Matsuki N, Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochem Biophys Res Commun* 1997; 237: 527-31.

- [19] Simpson RC, Freedland RA. Factors affecting the rate of gluconeogenesis from l-cysteine in the perfused rat liver. *J Nutr* 1976; 106: 1272-8.
- [20] Kredich NM, Foote LJ, Keenen BS. The stoichiometry and kinetics of the inducible cysteine desulfhydrase from *Salmonella typhimurium*. *J Biol Chem* 1973; 248: 6187-97.
- [21] Searcy DG, Lee SH. Sulfur reduction by human erythrocytes. *J Exp Zool* 1998; 282: 310-22.
- [22] Ashbaugh DG, Bigelow DB, Petty TL, Levine BE. Acute respiratory distress in adults. *Lancet* 1967; 2: 319-23.
- [23] Ware LB, Matthay MA. The acute respiratory distress syndrome. *N Engl J Med* 2000; 342: 1334-49.
- [24] Bernard GR, Artigas A, Brigham KL, et al. The American-European consensus conference on ARDS: definitions, mechanisms, relevant outcomes, and clinical trial coordination. *Am J Respir Crit Care Med* 1994; 149: 818-24.
- [25] Hudson LD, Milberg JA, Anardi D, Maunder RJ. Clinical risks for development of the acute respiratory distress syndrome. *Am J Respir Crit Care Med* 1995; 151: 293-301.
- [26] Krafft P, Fridrich P, Pernerstorfer T, et al. The acute respiratory distress syndrome: definitions, severity and clinical outcome. An analysis of 101 clinical investigations. *Intensive Care Med* 1996; 22: 519-29.
- [27] Doyle RL, Szaflarski N, Modin GW, Wiener-Kronish JP, Matthay MA. Identification of patients with acute lung injury. Predictors of mortality. *Am J Respir Crit Care Med* 1995; 152: 1818-24.
- [28] Rubenfeld GD, Caldwell E, Peabody E, et al. Incidence and outcomes of acute lung injury. *N Engl J Med* 2005; 353: 1685-93.
- [29] Windsor AC, Walsh CJ, Mullen PG, et al. Tumor necrosis factor-alpha blockade prevents neutrophil CD18 receptor upregulation and attenuates acute lung injury in porcine sepsis without inhibition of neutrophil oxygen radical generation. *J Clin Invest* 1993; 91: 1459-68.
- [30] Astiz ME, Rackow EC. Septic shock. *Lancet* 1998; 351: 1501-4.
- [31] Hui Y, Du J, Tang C, Bin G, Jiang H. Changes in arterial hydrogen sulfide (H<sub>2</sub>S) content during septic shock and endotoxin shock in rats. *J Infect* 2003; 47: 155-60.
- [32] Zhang H, Mochhala SM, Bhatia M. Endogenous hydrogen sulfide regulates inflammatory response in polymicrobial sepsis by activating the extracellular signal-regulated kinase (ERK)-NFκB pathway. *J Immunol* 2008; 181: 4320-31.
- [33] Bhatia M, Zhi L, Zhang H, Ng SW, Moore PK. Role of substance P in hydrogen sulfide-induced pulmonary inflammation in mice. *Am J Physiol Lung Cell Mol Physiol* 2006; 291: L896-904.
- [34] Tamizhselvi R, Moore PK, Bhatia M. Hydrogen sulfide acts as a mediator of inflammation in acute pancreatitis: *in vitro* studies using isolated mouse pancreatic acinar cells. *J Cell Mol Med* 2007; 11: 315-26.
- [35] Bhatia M, Wong FL, Fu D, Lau HY, Mochhala SM, Moore PK. Role of hydrogen sulfide in acute pancreatitis and associated lung injury. *FASEB J* 2005; 19: 623-5.
- [36] Tamizhselvi R, Moore PK, Bhatia M. Inhibition of hydrogen sulfide synthesis attenuates chemokine production and protects mice against acute pancreatitis and associated lung injury. *Pancreas* 2008; 36: e24-31.
- [37] Esecchie A, Kiss L, Olah G, et al. Protective effect of hydrogen sulfide in a murine model of combined burn and smoke inhalation-induced acute lung injury. *Clin Sci (Lond)* 2008; 115: 91-7.
- [38] Li T, Zhao B, Wang C, et al. Regulatory effects of hydrogen sulfide on IL-6, IL-8 and IL-10 levels in the plasma and pulmonary tissue of rats with acute lung injury. *Exp Biol Med (Maywood)* 2008; 233: 1081-7.
- [39] Bhatia M, Sidhapuriwala JN, Sparatore A, Moore PK. Treatment with H<sub>2</sub>S-releasing diclofenac protects mice against acute pancreatitis associated lung injury. *Shock* 2008; 29: 84-8.
- [40] Li L, Rossoni G, Sparatore A, Lee LC, Del Soldato P, Moore PK. Anti-inflammatory and gastrointestinal effects of a novel diclofenac derivative. *Free Radic Biol Med* 2007; 42: 706-19.
- [41] Beck-Speier I, Liese JG, Belohradsky BH, Godleski JJ. Sulfite stimulates NADPH oxidase of human neutrophils to produce active oxygen radicals via protein kinase C and Ca<sup>2+</sup>/calmodulin pathways. *Free Radic Biol Med* 1993; 14: 661-8.
- [42] Shigehara T, Mitsuhashi H, Ota F, et al. Sulfite induces adherence of polymorphonuclear neutrophils to immobilized fibrinogen through activation of Mac-1 β2-integrin (CD11b/CD18). *Life Sci* 2002; 70: 2225-32.
- [43] Mitsuhashi H, Yamashita S, Ikeuchi H, et al. Oxidative stress-dependent conversion of hydrogen sulfide to sulfite by activated neutrophils. *Shock* 2005; 24: 529-34.
- [44] Mitsuhashi H, Ikeuchi H, Yamashita S, et al. Increased levels of serum sulfite in patients with acute pneumonia. *Shock* 2004; 21: 99-102.
- [45] Zhi L, Ang AD, Zhang H, Moore PK, Bhatia M. Hydrogen sulfide induces the synthesis of proinflammatory cytokines in human monocyte cell line U937 via the ERK-NFκB pathway. *J Leukoc Biol* 2007; 81: 1322-32.
- [46] Rinaldi L, Gobbi G, Pambianco M, Micheloni C, Mirandola P, Vitale M. Hydrogen sulfide prevents apoptosis of human PMN via inhibition of p38 and caspase 3. *Lab Invest* 2006; 86: 391-7.
- [47] Oh GS, Pae HO, Lee BS, et al. Hydrogen sulfide inhibits nitric oxide production and nuclear factor-κB via heme oxygenase-1 expression in RAW264.7 macrophages stimulated with lipopolysaccharide. *Free Radic Biol Med* 2006; 41: 106-19.
- [48] Hu LF, Wong PT, Moore PK, Bian JS. Hydrogen sulfide attenuates lipopolysaccharide-induced inflammation by inhibition of p38 mitogen-activated protein kinase in microglia. *J Neurochem* 2007; 100: 1121-8.
- [49] Zhang H, Zhi L, Mochhala SM, Moore PK, Bhatia M. Endogenous hydrogen sulfide regulates leukocyte trafficking in cecal ligation and puncture-induced sepsis. *J Leukoc Biol* 2007; 82: 894-905.
- [50] Fiorucci S, Antonelli E, Distrutti E, et al. Inhibition of hydrogen sulfide generation contributes to gastric injury caused by non-steroidal anti-inflammatory drugs. *Gastroenterology* 2005; 129: 1210-24.
- [51] Zanoardo RC, Brancalone V, Distrutti E, Fiorucci S, Cirino G, Wallace JL. Hydrogen sulfide is an endogenous modulator of leukocyte-mediated inflammation. *FASEB J* 2006; 20: 2118-20.
- [52] Groneberg DA, Frossard QN, Fischer A. Neurogenic mechanisms in bronchial inflammatory disease. *Allergy* 2004; 59: 1139-52.
- [53] O'Connor TM, O'Connell J, O'Brien DI, Goode T, Bredin CP, Shanahan F. The role of substance P in inflammatory disease. *J Cell Physiol* 2004; 201: 167-80.
- [54] Bhatia M, Mochhala SM. Role of inflammatory mediators in the pathophysiology of acute respiratory distress syndrome. *J Pathol* 2004; 202: 145-56.
- [55] Trevisani M, Patacchini R, Nicoletti P, et al. Hydrogen sulfide causes vanilloid receptor 1-mediated neurogenic inflammation in the airways. *Br J Pharmacol* 2005; 145: 1123-31.
- [56] Zhang H, Hegde A, Ng SW, Adhikari S, Mochhala SM, Bhatia M. Hydrogen sulfide up-regulates substance P in polymicrobial sepsis-associated lung injury. *J Immunol* 2007; 179: 4153-60.
- [57] Bhatia M, Sidhapuriwala J, Ng SW, Tamizhselvi R, Mochhala S. Proinflammatory effects of hydrogen sulfide on substance P in caerulein-induced acute pancreatitis. *J Cell Mol Med* 2008; 12: 580-90.

Received: February 12, 2009

Revised: March 13, 2009

Accepted: March 17, 2009

© Zhang and Bhatia; licensee *Bentham Open*.This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.