

Antileukemic Properties and Structure-Activity Relationships of *O*- and *S*-Glycosylated Derivatives of Juglone and Related 1,4-Naphthoquinones

S.N. Fedorov*, L.K. Shubina, A.S. Kuzmich and S.G. Polonik

Pacific Institute of Bioorganic Chemistry, Vladivostok, 690022, Russian Federation

Abstract: Glycosylated derivatives of physiologically active natural compound juglone and related 1,4-naphthoquinones are known as antifungal, immunomodulatory, and antitumor substances. However, their antileukemic properties and structure-activity relationships have been studied insufficiently. Antileukemic effects and structure-activity relationships (SAR) of the 50 1,4-naphthoquinone derivatives were examined using HL-60 human promyelocytic leukemia cells and MTS method of the study of cell viability. As was shown, the substances inhibited viability of HL-60 cells at the wide range of concentrations. SAR study revealed the structure peculiarities which lead to increase or decrease of the antileukemic activity of the compounds studied. In conclusion, *O*- or *S*-glycosylated derivatives of juglone and related 1,4-naphthoquinones have potential for development of the new antileukemic agents and should be further investigated.

Keywords: Glycosides of 1,4-naphthoquinones, juglone, lawsone, phthiocol derivatives, antileukemic properties, SAR study.

INTRODUCTION

As early as in 1973 Cote and Goodman first reported the synthesis and isolation of four 1,4-naphthoquinone *O*-glycosides [1]. No biological activity for these compounds was published. In 1975-1977 a Brazilian scientific group published synthesis, structures, and biological activity of five other 1,4-naphthoquinone *O*-glycosides. The compounds were reported to be effective *in vivo* against rat tumor Walker 256 carcinosarcoma, mouse lymphocytic leukemia P-388, and Ehrlich ascitic tumor [2,3]. Since that time synthesis, isolation and cytotoxic, antitumor, immunomodulatory, and antifungal properties of *O*-glycosides of natural products juglone, lapachol, lawsone, shikonin, and related 1,4-naphthoquinones were reported [4-8].

In 1983 – 2010 years, we reported syntheses, structure elucidations, and biological activities of about 100 new *O*- or *S*-glycosides of 1,4-naphthoquinones and products of their intramolecular cyclisation [9-20]. Many of these compounds, in distinct from the earlier studied those, have a carbohydrate moiety attached to the quinone part of the molecule *via* sulfur atom (thioglycosyl group). The synthesized glycosides were shown to possess earlier known kinds of activity and some of them demonstrated new one, the induction of expression of heat-shock protein Hsp70 [21].

However, antileukemic properties and structure-activity relationships (SAR) of the glycosylated derivatives of 1,4-naphthoquinones have been studied insufficiently. In the present paper, antileukemic effects and SAR of the 50 of 1,4-naphthoquinone derivatives were examined using HL-60 human promyelocytic leukemia cells and MTS method of the study of cell viability.

MATERIALS AND METHODOLOGY

Drugs and Chemicals

The systematic names of the compounds studied **1-50** are shown in Supplementary Material. These compounds were synthesized and purified as described previously [9-20] and were pure in accordance with chromatographic and NMR data. RPMI medium was from Gibco Invitrogen Corporation (Carlsbad, CA, USA). Fetal bovine serum (FBS) was from Gemini Bio-Products (Calabasas, CA, USA); penicillin and streptomycin were from Bio-Whittaker (Walkersville, MD, USA); L-glutamine was from Mediatech, Inc. (Herndon, Virginia, USA). The MTS (5-(3-carboxymethoxyphenyl)-2-(4,5-dimethylthiazolyl)-3-(4-sulfophenyl) tetrazolium, inner salt) reagent kit for the cell proliferation assay was from Promega (Madison, WI, USA).

Cell Culture

The human promyelocytic leukemia HL-60 cell line was obtained from the American Type Culture Collection (Rockville, MD, USA) and were cultured at 37°C and 5% CO₂ in RPMI medium containing 10% FBS, 2 mM L-glutamine, 100 units/ml penicillin and 100 µg/ml streptomycin. Information regarding the genetic background of HL-60 cell line is available online.

Cell Viability Assay

The effect of the glycosides on cell viability was evaluated using MTS reduction into its formazan product [22]. The HL-60 cells were cultured for 12 h in 96-well plates (6,000 cells/well in 50 µl of medium). Then 50 µl of medium containing glycosides at various concentrations were added and the cells were incubated for 22 h. Then 20 µl of the MTS reagent were added into each well and MTS reduction was measured 2 h later spectrophotometrically at 492 nm and 690 nm as background using the Multiskan MS microplate reader (Labsystems, Finland). Results are shown in Table 3 and represent the IC₅₀ of the substances against HL-60 cells.

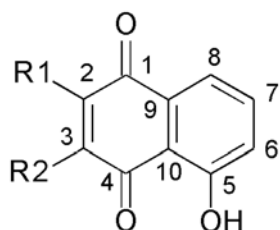
*Address correspondence to this author at the Pacific Institute of Bioorganic Chemistry, 159 Prospect 100-let Vladivostoku, Vladivostok, 690022, Russian Federation; Tel: 7(4232)311168; Fax: 7(4232)314050; E-mail: fedorov@piboc.dvo.ru

Statistics

The computer programs, Excel and Statistica 6.0 for Windows (StatSoft, Inc., Tulsa, OK, USA, 2001) were used for calculation of IC₅₀ and other analyses of the obtained data.

RESULTS

We divided compounds into three structural groups: first, *O*- and *S*- glycosides of juglone (5-hydroxy-1,4-naphthoquinone) (**1-16**, Table 1); second, *O*- and *S*- glycosides of other 1,4-naphthoquinones (**17-40**, Table 2); third, products of intramolecular cyclisation of the *S*-glycosides of 1,4-naphthoquinones (**41-50**).



Juglone, R1 = R2 = H

The structures of the sugar parts of the substances (**1-40**) from Tables 1 and 2 are given below the Table 2:

Antileukemic effects of the compounds **1-50** and their structure-activity relationships were examined using HL-60

- Two sugar residues in the molecule significantly reduce activity of the substances (compare **18, 19** with **32, 20-24** or **3** with **1, 2** or **8** with **7**). Furthermore, two -OAc groups at positions 5, 8 of the quinone moiety, renew the activity (compare **17** with **18, 19**).
- The presence of a disaccharide residue in the molecule also significantly reduces activity of compounds (**25, 26, 50**) as compared to compounds with a monosaccharide residue in the molecule (**20-24, 32, 41-49**);
- Juglone derivatives possessing acetylated sugar moiety (**1-14**) are significantly more active than that possessing sugar moiety with free hydroxyl groups (**15, 16**).
- Juglone derivatives possessing various sugar moieties in the molecules (**1, 2, 4, 5, 7, 10-14**) show almost equal activities.

DISCUSSION

Glycosides of 1,4-naphthoquinones are synthetically available biologically active compounds. Glycosylated derivatives of the natural compounds juglone, lawsone, phthiocol, shikonin, and echinochrome are among them. These compounds possess antitumor, immunomodulatory, antifungal and cytotoxic activities. Furthermore, lately some details of their mechanism of action became more pronounced, when their ability to induce the expression of heat-shock protein Hsp70 has been published [21]. Heat shock proteins are stress proteins and their upregulation is described as part

Table 1. Structures of the Juglone Glycosides 1-16

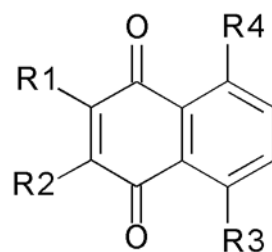
Glycoside number	Radicals	Glycoside number	Radicals
1	R ₁ = Ac ₄ βD-GlcO; R ₂ = H	9	R ₁ = Ac ₄ βD-GlcS; R ₂ = CH ₃
2	R ₁ = H; R ₂ = Ac ₄ βD-GlcO	10	R ₁ = H; R ₂ = Ac ₄ βD-GalS
3	R ₁ = R ₂ = Ac ₄ βD-GlcO	11	R ₁ = H; R ₂ = Ac ₃ βD-XylS
4	R ₁ = Ac ₇ βD-MalO; R ₂ = H	12	R ₁ = H; R ₂ = Ac ₃ αL-AraS
5	R ₁ = H; R ₂ = Ac ₇ βD-MalO	13	R ₁ = H; R ₂ = Ac ₇ βD-MalS
6	R ₁ = Ac ₄ βD-GlcS; R ₂ = H	14	R ₁ = H; R ₂ = Ac ₄ βD-ManS
7	R ₁ = H; R ₂ = Ac ₄ βD-GlcS	15	R ₁ = βD-GlcS; R ₂ = CH ₃
8	R ₁ = R ₂ = Ac ₄ βD-GlcS	16	R ₁ = βD-GlcS; R ₂ = CH ₂ CH ₂ CH ₃

human promyelocytic leukemia cells and MTS method of the study of cell viability. The results are shown in Table 3 and represent the IC₅₀ of the substances against HL-60 cells.

In accordance with the data presented in Table 3 the following structure-activity relationships for compounds **1-50** can be concluded:

- The most active compounds are glycosides of juglone **1-14** and products of intramolecular cyclisation of the *S*-glycosides of 1,4-naphthoquinones **41-49**;
- Electronegative groups like -OH, -Cl, -NH₂, -OMe, -OAc at the α-position to the sugar moiety dramatically reduce antileukemic activity of the substances **29-31, 33-40**;

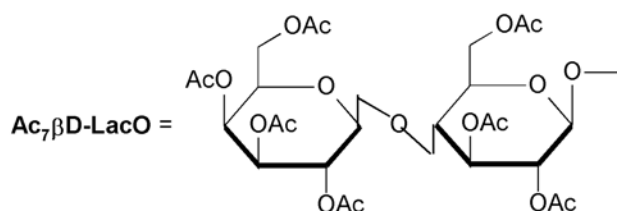
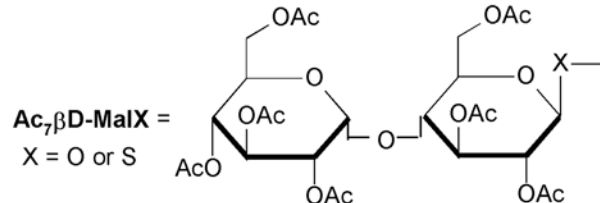
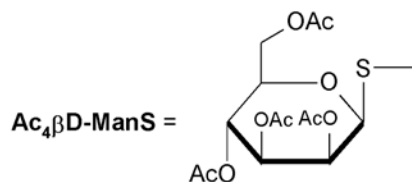
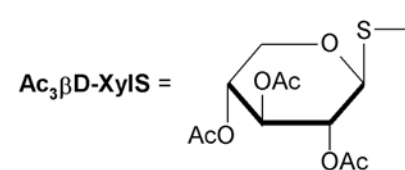
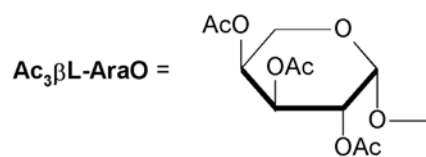
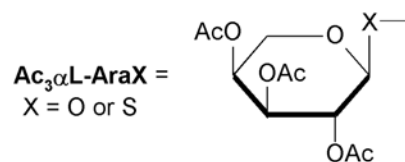
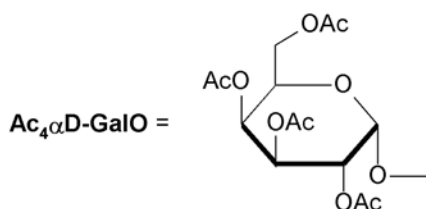
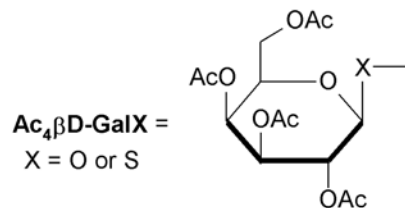
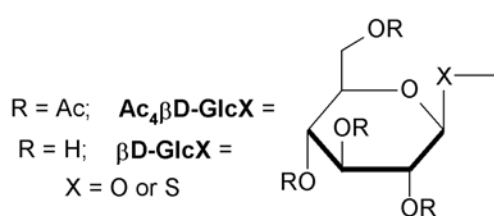
of the stress response. Production of high levels of heat shock proteins can be triggered by infection, inflammation,



1,4-Naphthoquinone,
R1 = R2 = R3 = R4 = H

Table 2. Structures of the 1,4-Naphthoquinone Glycosides 17-40

Subst. #	Radicals	Subst. #	Radicals
17	$R_1=R_2=Ac_4\beta D-GlcS; R_3=R_4=OAc$	29	$R_1=Ac_4\beta D-GlcO; R_2=OH; R_3=R_4=H$
18	$R_1=R_2=Ac_4\beta D-GlcS; R_3=R_4=H$	30	$R_1=Ac_4\beta D-GlcO; R_2=OMe; R_3=R_4=H$
19	$R_1=R_2=Ac_3\beta D-XylS; R_3=R_4=H$	31	$R_1=\beta D-GlcO; R_2=OH; R_3=R_4=H$
20	$R_1=Ac_4\beta D-GlcO; R_2=R_3=R_4=H$	32	$R_1=Ac_4\beta D-GlcS; R_2=R_3=R_4=H$
21	$R_1=Ac_4\beta D-GalO; R_2=R_3=R_4=H$	33	$R_1=Ac_4\beta D-GlcS; R_2=Cl; R_3=R_4=H$
22	$R_1=Ac_4\alpha D-GalO; R_2=R_3=R_4=H$	34	$R_1=Ac_4\beta D-GlcS; R_2=OAc; R_3=R_4=H$
23	$R_1=Ac_3\alpha L-AraO; R_2=R_3=R_4=H$	35	$R_1=Ac_4\beta D-GlcS; R_2=OH; R_3=R_4=H$
24	$R_1=Ac_3\beta L-AraO; R_2=R_3=R_4=H$	36	$R_1=Ac_4\beta D-GlcS; R_2=NH_2; R_3=R_4=H$
25	$R_1=Ac_7\beta D-LacO; R_2=R_3=R_4=H$	37	$R_1=Ac_4\beta D-GlcS; R_2=OMe; R_3=R_4=H$
26	$R_1=Ac_7\beta D-MalO; R_2=CH_3; R_3=R_4=H$	38	$R_1=Ac_3\beta D-XylS; R_2=OMe; R_3=R_4=H$
27	$R_1=Ac_4\beta D-GlcO; R_2=CH_3; R_3=R_4=H$	39	$R_1=Ac_4\beta D-ManS; R_2=OMe; R_3=R_4=H$
28	$R_1=Ac_4\beta D-GlcO; R_2=Et; R_3=R_4=H$	40	$R_1=Ac_3\alpha L-AraS; R_2=OMe; R_3=R_4=H$



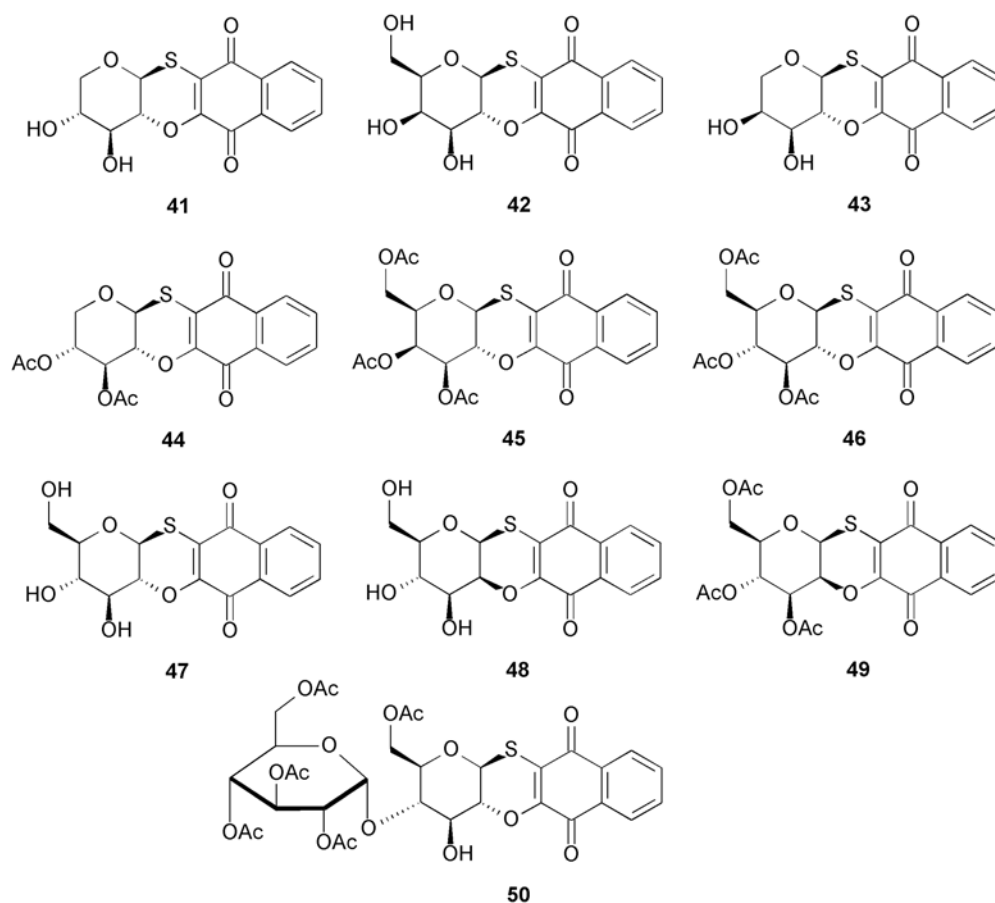


Table 3. IC₅₀ of 1,4-Naphthoquinone Glycosides 1-50 Against HL-60 Cells

Subst. #	IC ₅₀ , μM	Subst. #	IC ₅₀ , μM	Subst. #	IC ₅₀ , μM	Subst. #	IC ₅₀ , μM
1	0.6	14	1.4	27	8.0	40	> 20.0*
2	0.7	15	13.3	28	8.9	41	1.0
3	2.4	16	10.0	29	> 100.0*	42	1.1
4	1.1	17	6.7	30	15.0	43	1.6
5	1.0	18	17.6	31	> 200.0*	44	1.6
6	2.3	19	18.8	32	3.5	45	1.8
7	1.3	20	7.0	33	48.0	46	1.5
8	4.8	21	6.9	34	> 20.0*	47	1.1
9	0.6	22	3.4	35	> 100.0*	48	5.0
10	1.6	23	4.6	36	22.8	49	1.5
11	1.1	24	8.8	37	20.2	50	> 20.0*
12	1.2	25	24.7	38	> 20.0*		
13	1.0	26	15.7	39	> 20.0*		

*Maximal dose studied.

and, in particular, by exposure to the cell toxins [23]. These proteins have a significant role in cardiovascular diseases, immunity, and carcinogenesis [24-26]. Thus, it was important to study SAR of a big row of 1,4-naphthoquinone gly-

cosides in order to understand how to synthesize a compound with the desirable properties.

SAR study showed the juglone derivatives were among the most active compounds. This finding demonstrated the

importance of the hydroxyl group at the position 5 of the quinone moiety for the increase of the activity. Another one confirmation of this supposition was the fact that compound **17** possessing -OAc groups at the positions 5 and 8 was much more active than compounds **18** and **19**. Compounds **29-31** and **33-40** possessing -OH, -Cl, -NH₂, -OMe, or -OAc groups at the α -position to the sugar moiety showed dramatically reduced antileukemic activity. Presence of two monosaccharide moieties in the molecule also significantly reduced the activity of compounds **18**, **19**. These findings may be explained by the known effect of stabilization of the glycoside bond in the presence of -OH or relative groups [12] that in turn may lead to the stability of similar compounds inside the cells. We also concluded that the antileukemic activity of compounds does not depend on the kind of a monosaccharide residue presenting in the structures of the substances studied.

The search for glycosides of 1,4-naphthoquinones that possess more high antitumor, immunomodulatory, antifungal and Hsp70 upregulative activities will create opportunities for selection of new anticancer agents. We hope the revealed SAR can help to synthesize 1,4-naphthoquinone glycosides with needful activities.

CONCLUSION

In conclusion, *O*- or *S*- Glycosylated derivatives of juglone and related 1,4-naphthoquinones have potential for development of the new antileukemic agents and should be further investigated.

ACKNOWLEDGEMENTS

This work was supported by the Grant NSS 3531.2010.4 from the President of RF, Program of Presidium of RAS "Molecular and Cell Biology", and FEB RAS Grant 09-III-A-05-146.

SUPPLEMENTARY MATERIAL

Supplementary material is available on the publishers Web site along with the published article.

REFERENCES

- [1] Cote PN, Goodman L. Glucopyranosides derived from 2-hydroxy-1,4-naphthoquinones. *Carbohydr Res* 1973; 26: 247-51.
- [2] Linardi MCF, de Oliveira MM, Sampaio MRP. A lapachol derivative active against mouse lymphocytic leukemia P-388. *J Med Chem* 1975; 18: 1159-61.
- [3] de Oliveira MM, Linardi MCF, Sampaio MRP. Effects of quinone derivatives on an experimental tumor. *J Pharm Sci* 1977; 66: 562-3.
- [4] Steinerova N, Cudlin J, Vanek Z. Glucosidation of 2-hydroxy- and 5-hydroxy-1,4-naphthoquinone. *Collection Czechoslov Chem Commun* 1980; 45: 2684-7.
- [5] Sterzl J, Cudlin J, Steinerova N, Johanovska D, Milerova J. The immunoinhibitory and immunostimulatory effects of hydroxyanthra- and hydroxynaphthoquinone derivatives. *Folia Microbiol* 1981; 26: 169-75.
- [6] Otten SL, Rosazza JP. Microbial transformations of natural antitumor agents. 17. Conversions of lapachol by *Cunninghamella echinulata*. *J Nat Prod* 1981; 44: 562-8.

- [7] Ozgen U, Kazaz C, Secen H, Calis I, Coskun M, Houghton PJ. A novel naphthoquinone glycoside from *Rubia peregrina* L. *Turk J Chem* 2009; 33: 561-8.
- [8] Su Y, Xie J, Wang Y, Hu X, Lin X. Synthesis and antitumor activities of new shikonin glycosides. *Eur J Med Chem* 2010; 45: 2713-8.
- [9] Polonik SG, Tolkach AM, Uvarova NI. Synthesis of acetylated glycosides of hydroxynaphthoquinones. *Chem Nat Comp* 1983; 19: 307-10.
- [10] Polonik SG, Tolkach AM, Denisenko VA, Uvarova NI. Synthesis of glucosides of 3-alk[enyl]-2-hydroxy-1,4-naphthoquinones. *Chem Nat Comp* 1983; 19: 310-3.
- [11] Polonik SG, Tolkach AM, Uvarova NI, Stekhova SI, Shentsova EB, Anisimov MM. Synthesis and anti-fungal activity of acetylated glycosides of 1,4-naphthoquinone. *Pharm Chem J* 1986; 20: 93-7.
- [12] Polonik SG, Tolkach AM, Uvarova NI. Autocatalytic condensation of 1,2-orthoesters of sugars with 2,3-dihydroxy-1,4-naphthoquinone (isonaphthazarin). *Chem Nat Comp* 1989; 25: 406-10.
- [13] Tolkach AM, Polonik SG, Stekhova SI, Prokofieva NG, Uvarova NI. Synthesis, cytotoxic and antifungal activity of acetylated thioglycosides of 1,4-naphthoquinones. *Pharm Chem J* 1989; 23: 1013-6.
- [14] Polonik SG, Tolkach AM, Stekhova SI, Shentsova EB, Uvarova NI. Synthesis of acetylated glycosides of hydroxyjuglones and study of their antifungal activity. *Pharm Chem J* 1992; 26: 500-2.
- [15] Polonik SG, Tolkach AM, Shentsova EB, Uvarova NI. Synthesis and cytostatic activity of 2-bromo-3-alkyljuglones and related thioglycosides derived from them. *Pharm Chem J* 1995; 29: 668-71.
- [16] Polonik SG, Tolkach AM, and Uvarova NI. Reaction of acetylated 1,4-naphthoquinone thioglycosides with nucleophilic reagents. *Rus Chem Bul* 1996; 45: 459-61.
- [17] Polonik SG, Tolkach AM, Uvarova NI. Reaction of substituted 2,3-dichloronaphthazarines with acetylthioglucose. Synthesis of mono-, di-, tri-, and tetra- thioglycosides of naphthazarines. *Rus J Org Chem* 1998; 34: 1172-7.
- [18] Polonik SG, Prokofieva NG, Agafonova IG, Uvarova NI. Antitumor and immunostimulating activity of 5-hydroxy-1,4-naphthoquinone (juglone) *O*- and *S*-acetylglucosides. *Pharm Chem J* 2003; 37: 397-8.
- [19] Polonik SG. Synthesis and properties of water-soluble 5,8-dihydroxy-1,4-naphthoquinone thioglycosides structurally related to echinochrome. *Rus J Org Chem* 2009; 45: 1474-80.
- [20] Polonik SG. Glycosilation of shikonin by the Helferich method. *Chem Nat Comp* 2009; 45: 247-8.
- [21] Eremente EM, Antimonova OI, Shekalova OG, Polonik SG, Margulis BA, Guzhova IV. Novel compounds that increase expression of Hsp70 and its biological activity. *Cell Tiss Biol* 2010; 4: 251-7.
- [22] Baltrop JA, Owen TC, Cory AH, Cory JG. 5-(3-Carboxymethoxyphenyl)-2-(4,5-dimethylthiazolyl)-3-(4-sulfophenyl) tetrazolium, inner salt (MTS) and related analogs of 3-(4,5-dimethylthiazolyl)-2,5-diphenyltetrazolium bromide (MTT) reducing to purple water-soluble formazans as cell-viability indicators. *Bioorg Med Chem Lett* 1991; 1: 611-4.
- [23] Santoro MG. Heat shock factors and the control of the stress response. *Biochem Pharmacol* 2000; 59: 55-63.
- [24] Benjamin IJ, Mc Millan DR. Stress (heat shock) proteins: molecular chaperones in cardiovascular biology and disease. *Circulation Res* 1998; 83:117-32.
- [25] Nishikawa M, Takemoto S, Takakura Y. Heat shock protein derivatives for delivery of antigens to antigen presenting cells. *Int J Pharm* 2008; 354: 23-7.
- [26] Dai C, Whitesell L, Rogers AB, Lindquist S. Heat shock factor 1 is a powerful multifaceted modifier of carcinogenesis. *Cell* 2007; 130: 1005-18.